



FALCON-Phase: Integrating PacBio and Hi-C data for phased diploid genomes

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SFAF, May 23rd 2018, Santa Fe, NM

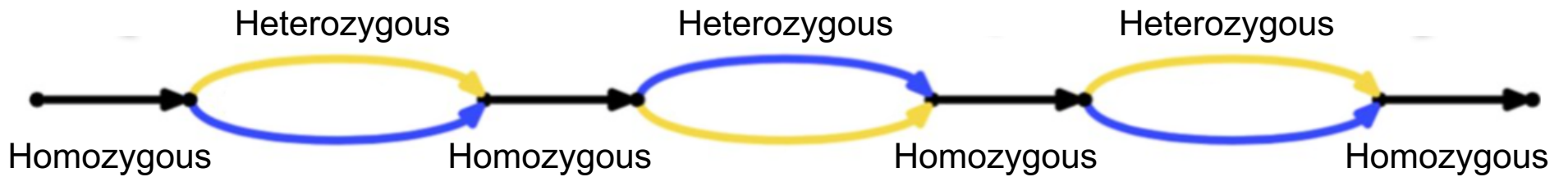
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Image Credit: Iannuzzi, 1996

HAPLOTYPE ASSEMBLY IN DIPLOIDS REMAINS A CHALLENGE

- Traditional assembly paradigm: choose inbred sample
- Non-model genome assembly is increasingly common
- Base accuracy and contiguity suffer if haplotypes collapsed

HAPLOTYPE ASSEMBLY IN DIPLOIDS REMAINS A CHALLENGE



“collapsed” haplotypes



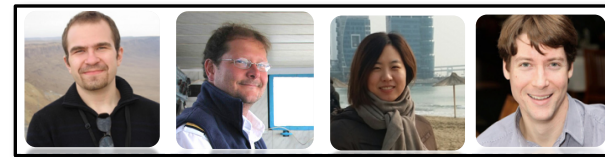
contiguous phased haplotigs



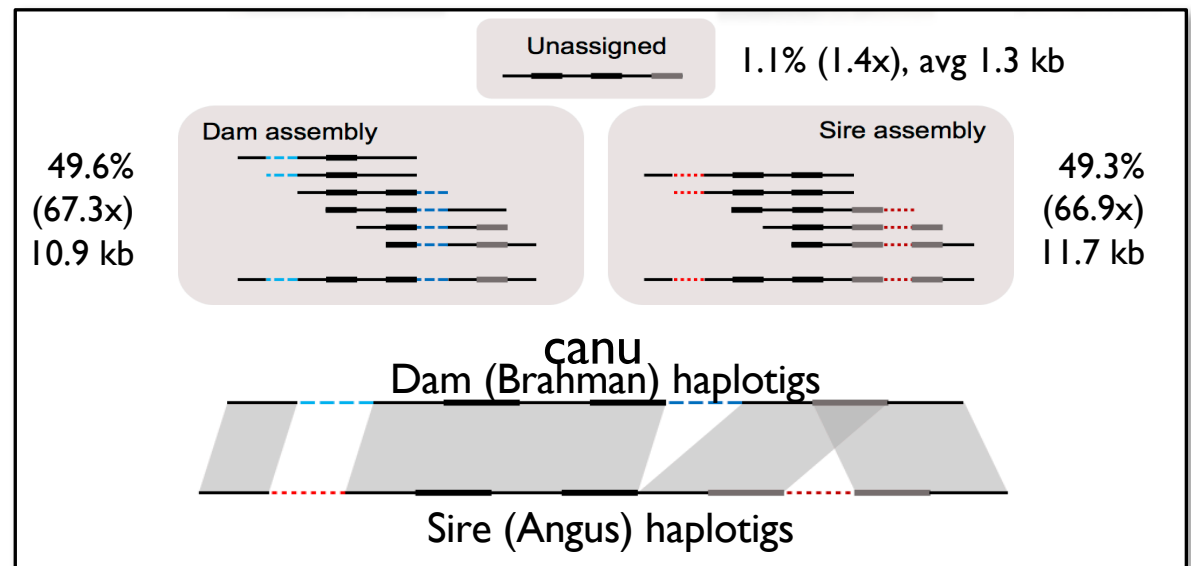
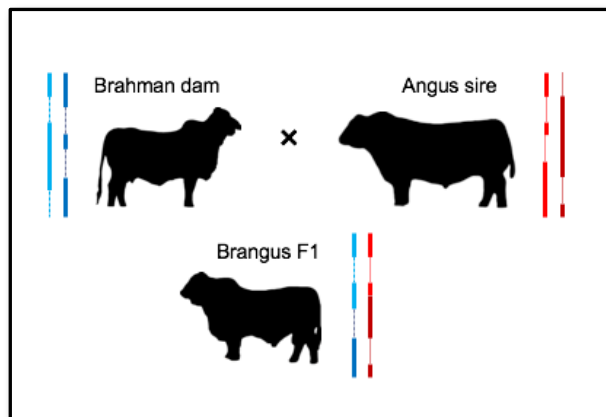
CURRENT APPROACHES TO LONG READ DIPLOID ASSEMBLY

1. Separate Reads with Trio Binning (**TrioCanu**)

- PacBio data for F1
- ILMN data for parent-specific k-mers
- Bin PacBio reads with k-mers
- Perform two haploid Canu assemblies



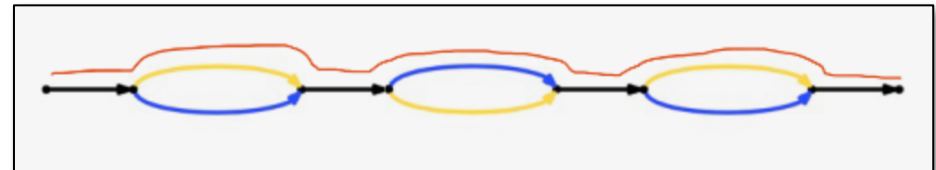
EXAMPLE ON F1 BULL



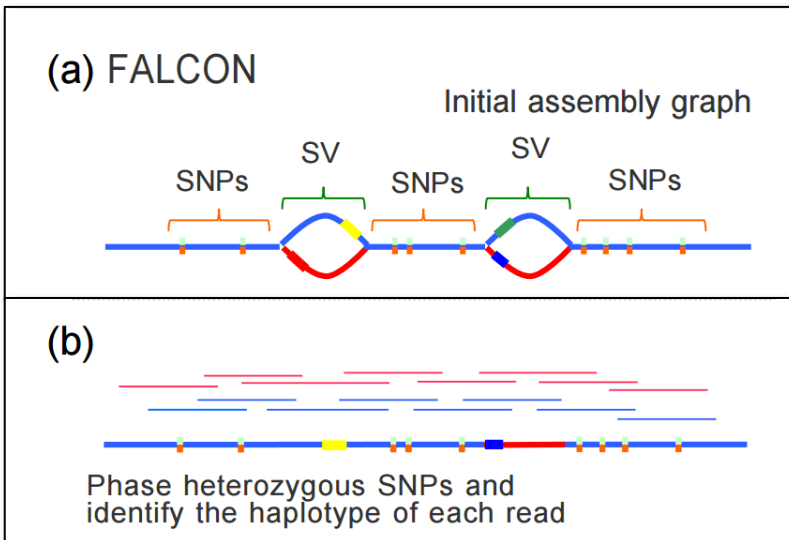
CURRENT APPROACHES TO LONG READ DIPLOID ASSEMBLY

2. Separate Haplotypes During Assembly with FALCON-Unzip

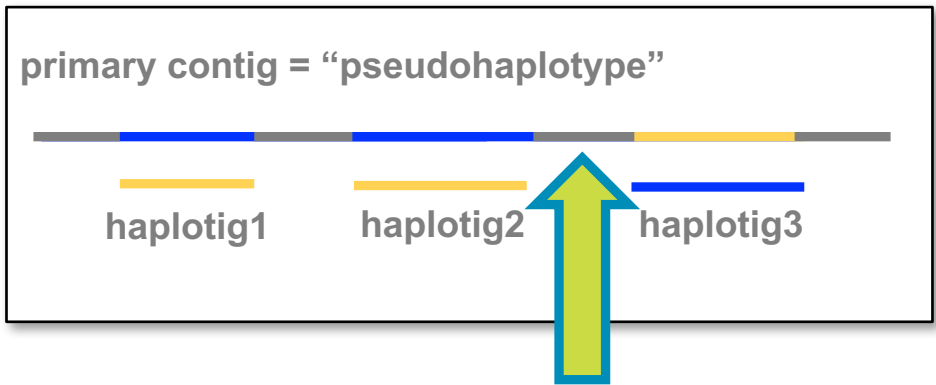
- PacBio data for diploid individual (no trio)
- Phase PacBio reads using SNPs identified in initial assembly graph
- Output phased and collapsed regions in high contiguity contigs



Weisenfeld et al. 2017



PSEUDOHAPLOTYPE AND HAPLOTIGS



"Phase/Haplotype Switch"

FALCON-PHASE: MOTIVATIONS AND GOALS

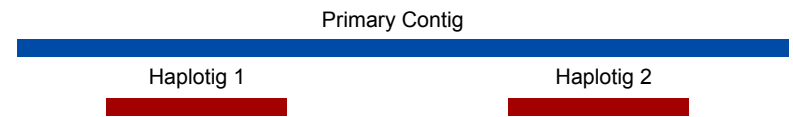
- FALCON-Unzip phase blocks are small
 - Phasing is function of heterozygosity, read depth, read length
 - Phase switches between haplotype blocks are nearly random
- Haplotype/phase switches are problematic
 - “Franken-haplotypes” impact base accuracy, gene prediction
 - Scaffolding errors
- Hi-C contains long-range haplotype information
- FALCON-Phase Tool
 - Open-source snakemake pipeline
 - Co-development project between PacBio and Phase Genomics
 - **Can be applied at contig and scaffold scale**



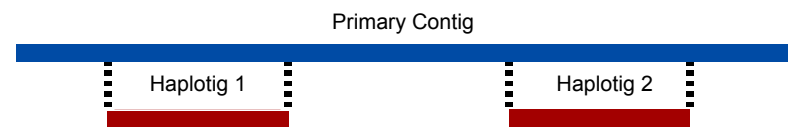
Zev Kronenberg

FALCON-PHASE WORKFLOW

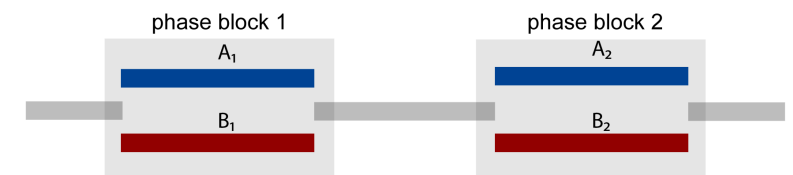
Input: FALCON-Unzip assembly & HiC data



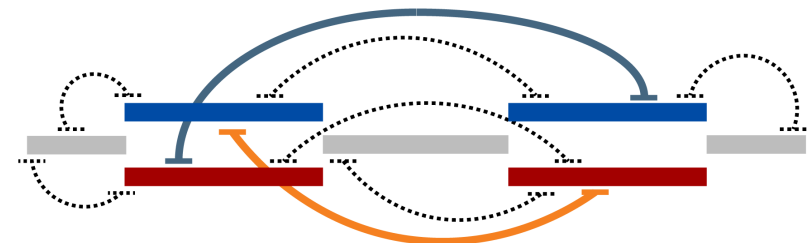
1. Identify **haplotig placement** on primary contigs



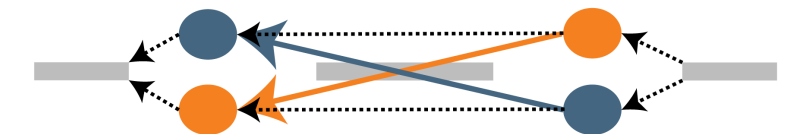
2. **Mince** primary contigs: separate **haplotig pairs** and **collapsed haplotypes**



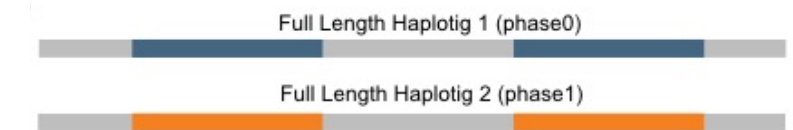
3. **Map** paired HiC reads to minced contigs and generate normalized **contact matrix**



4. **Phase** haplotigs along each primary contig



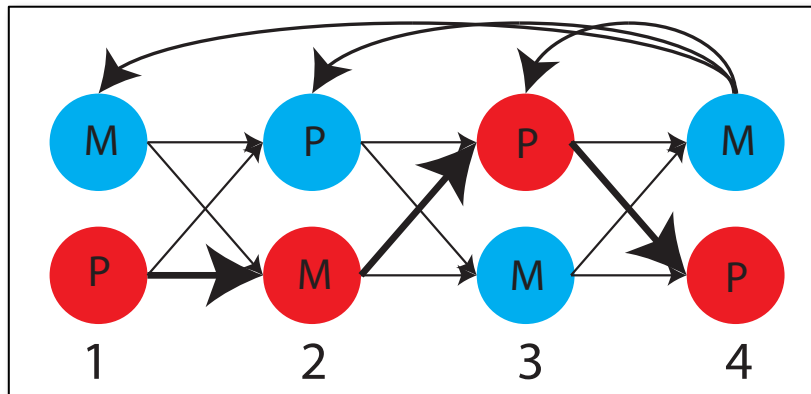
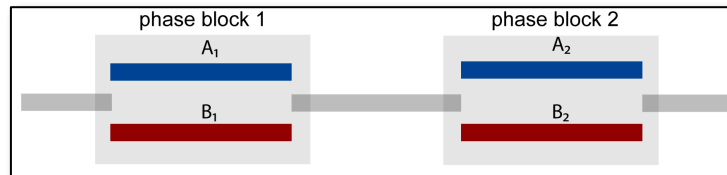
Output: phased full length haplotigs



PHASING ALGORITHM: INPUTS AND OUTPUTS

— FALCON-Unzip Input

- Order and pairing of phase blocks along primary contig



— Hi-C Input

- Normalized contact matrix between each phase block

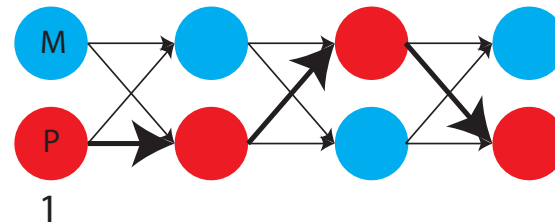
	A ₁	B ₁	A ₂	B ₂
A ₁	18	.	.	.
B ₁	1	15	.	.
A ₂	2	9	15	.
B ₂	7	0	3	12

— Output

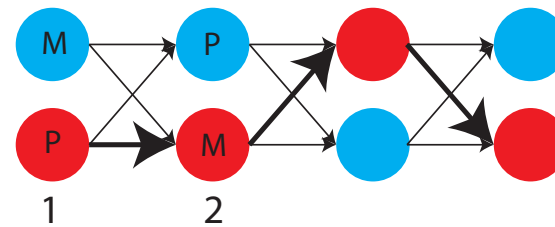
- Majority phase assignment configuration for haplotigs along primary contig

PHASING ALGORITHM

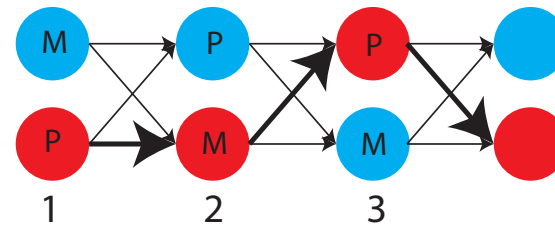
- Algorithm sweeps along phase-blocks of primary contig
- Phase assignment is conditioned on **all** those before and Hi-C links
- Process repeated for $> 10^7$ iterations (burn-in = 5×10^6)
- Complexity $\sim O(n^2)$
 - n phase blocks



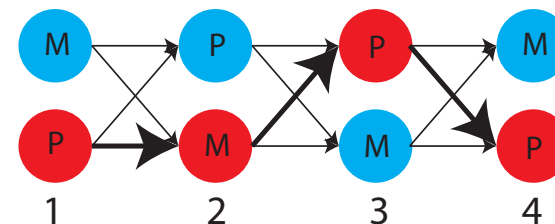
F(1)



F(2 | 1)

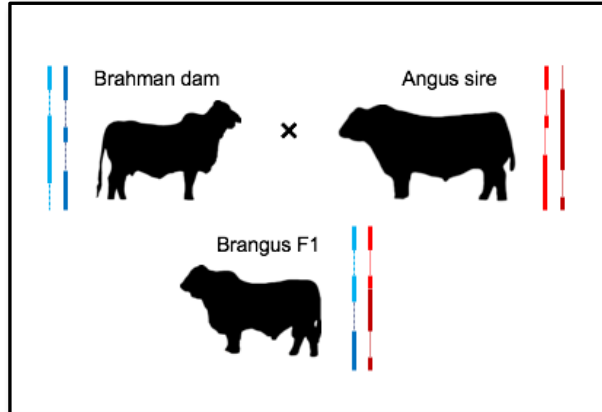


F(3 | 2, 1)



F(4 | 3, 2, 1)

VALIDATION DATASET: ANGUS-BRAHMAN F1 BULL



Data: Tim Smith (USDA), John Williams and Stefan Hiendleder (U Adelaide)

Canu Asms: Adam Phillippy, Sergey Koren, Arang Rhie (NHGRI)

FALCON-Unzip: 90% Unzipped

CONTIGS	NUMBER	LENGTH	N50
PRIMARY	1427	2.71 Gb	31.4 Mb
HAPLOTIGS	5879	2.45 Gb	2.48 Mb

Phase Genomics Hi-C

- 200 million read pairs

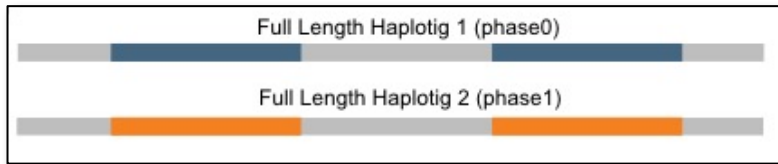
TrioCanu Assemblies

CONTIGS	NUMBER	LENGTH	N50
ANGUS DAM	1747	2.57 Gb	26.7 Mb
BRAHMAN SIRE	1040	2.68 Gb	23.3 Mb

Parental SNP Calls

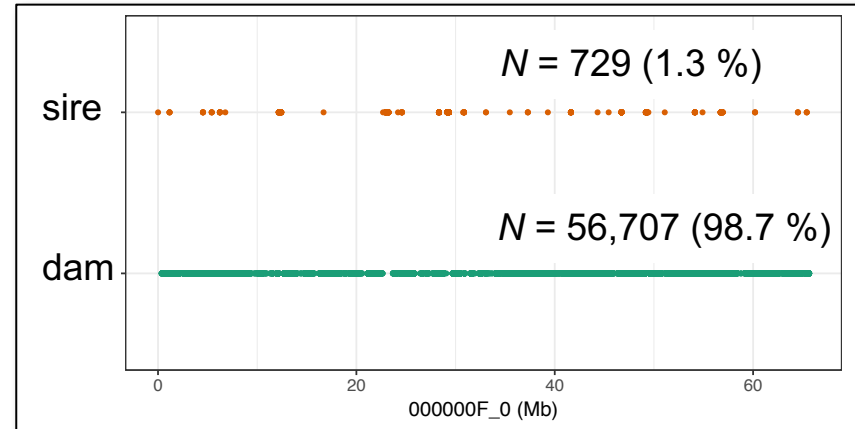
- 20-25x coverage ILM PE
- read mapping with bwa mem
- SNV calls with freebayes

PHASE ASSIGNMENT ACCURACY: PARENTAL SNV CALLS

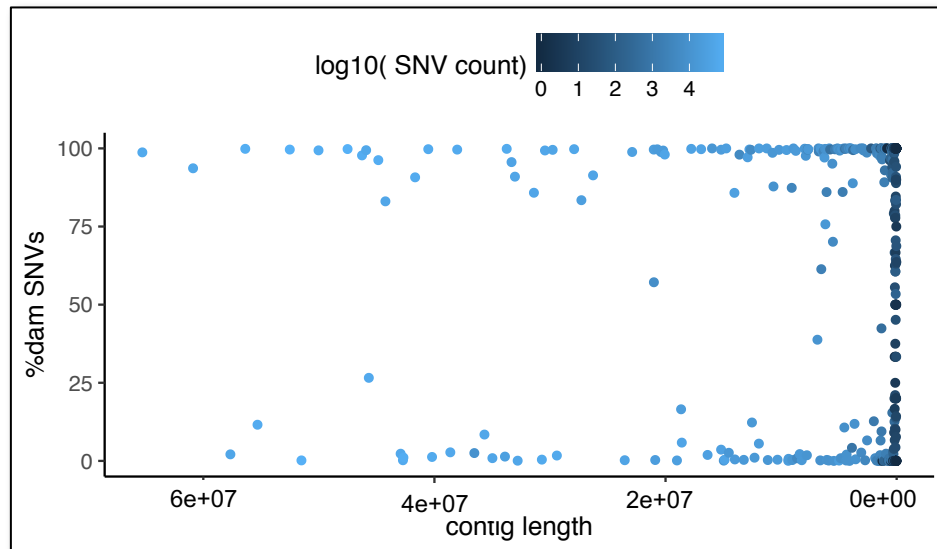


- bwa mem alignment of parental PE ILM data to phase0 haplotigs
- Variant calling with Freebayes
- SNV filtered for homozygous sites that differ between parents

PRIMARY CONTIG 000000F_0 (DAM)



ACCURACY BY PRIMARY CONTIG LENGTH

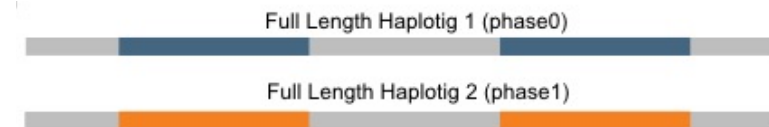


OVERALL PERFORMANCE

	SNV Count	Correct
dam	2,031,334	97.4 %
sire	1,464,748	95.8 %
total	3,496,082	96.7 %

PHASING CHROMOSOME-SCALE SCAFFOLDS

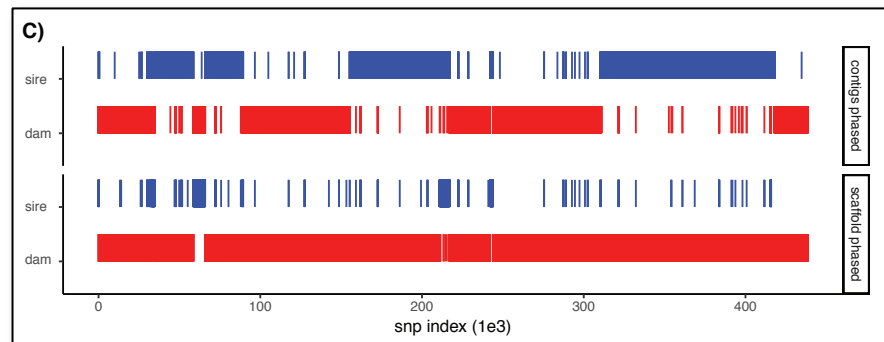
- Scaffold phase0 full-length haplotig with Proximo (Phase Genomics)
- Scaffolds are chromosome-scale
- We know:
 - order of contigs along scaffold
 - pairing of phase 0 and phase 1
- FALCON-Phase Scaffolds



Scaffold Phase 0 Haplotigs



PARENTAL SNVS AFTER SCAFFOLD PHASING



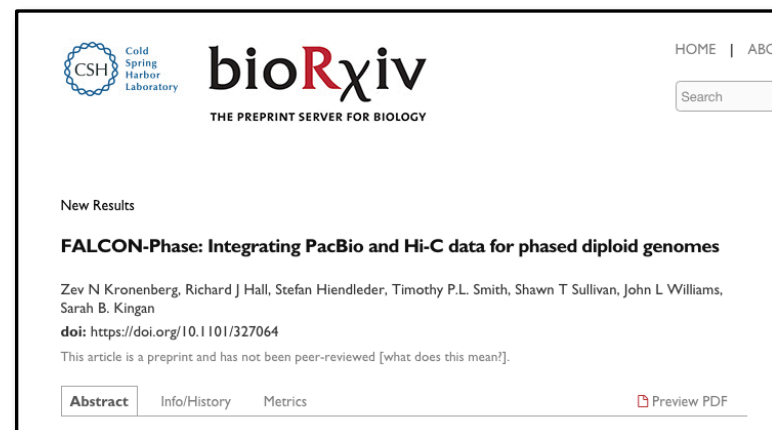
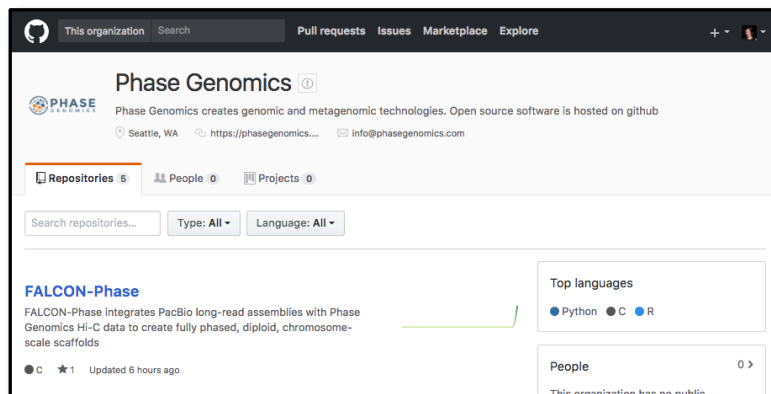
FALCON-Phase Scaffolds



Output: Chromosome-scale, phased, diploid assembly!

SUMMARY

- FALCON-Phase is highly accurate
 - > 96% accuracy when tested against parental assemblies or SNVs
- FALCON-Phase implemented in snakemake pipeline
 - Run locally or on cluster, Open source
- PacBio plus HiC is all you need to produce gapless, phased, chromosome-scale diploid assembly
- More Info:



ACKNOWLEDGEMENTS

—Coauthors

- Zev Kronenberg (Phase)
- Richard Hall (PacBio)
- Stefan Hiendleder (U Adelaide)
- Timothy Smith (USDA)
- Shawn Sullivan (Phase)
- John Williams (U Adelaide)



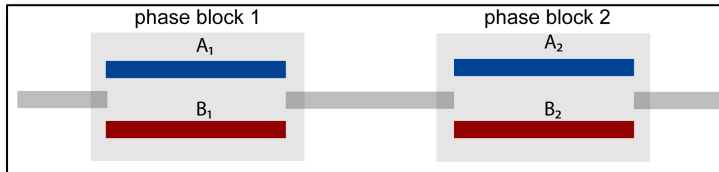
Greg Concepcion
Jonas Korlach
Billy Rowell
Ivan Sovic
Liz Tseng
Michelle Vierra



Ivan Liachko
Kaylee Mueller
Max Press
Andrew Wiser

Jason Chin
Mark Chaisson
Luke Harmon
Ryan Layer

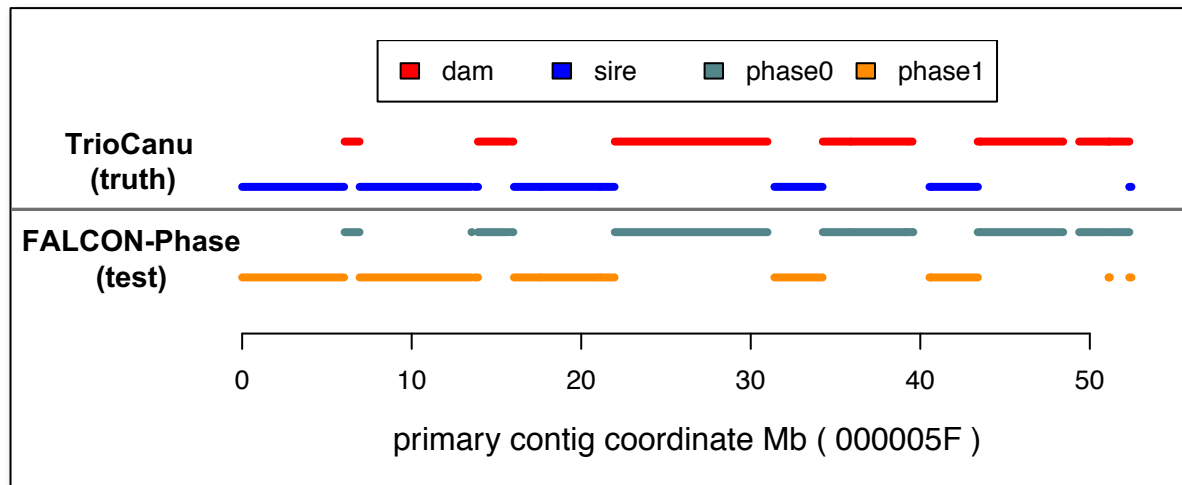
PHASE ASSIGNMENT ACCURACY: PARENTAL ASSIGNMENT



- Minimap to Canu Asms
- Highest PID for longest alignment
- Required concordance between pairs

Contig Assignment	Count	Length (%)	Mean Length
Dam	2,305	2.32 Gb (42 %)	1.01 Mb
Sire	2,305	2.32 Gb (42 %)	1.01 Mb
No Parent	1,704	116 Mb (2.1 %)	68.1 kb
Collapsed	3,934	374 Mb (6.8 %)	88.2 kb

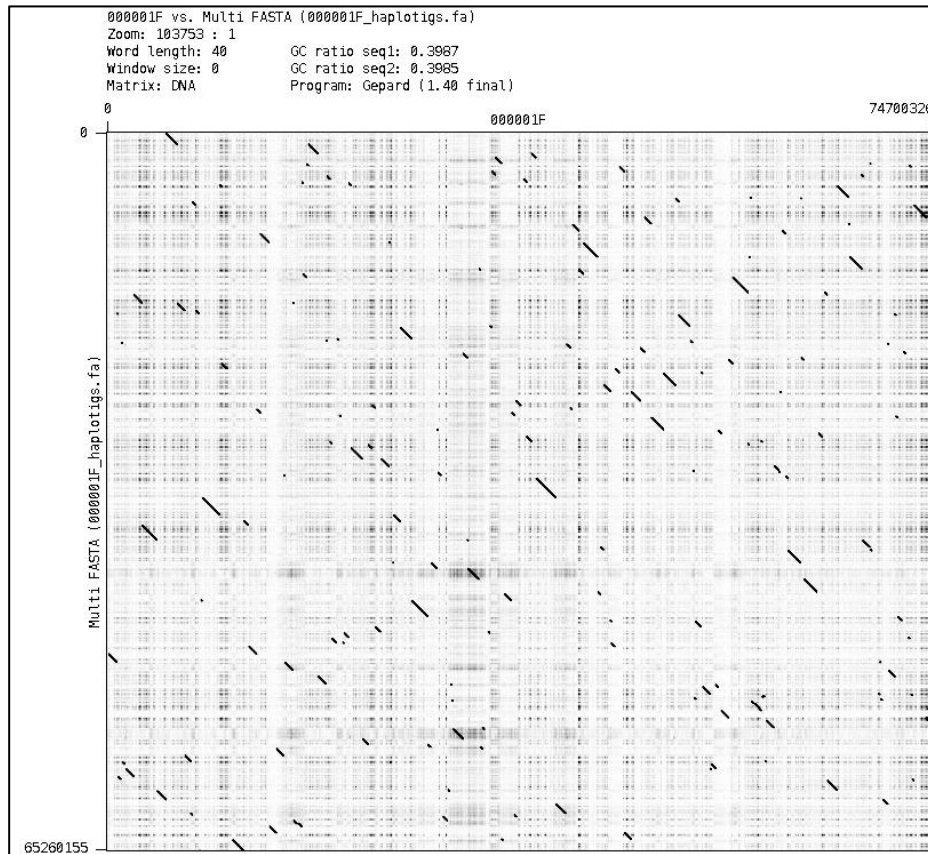
RESULTS FOR PRIMARY CONTIG 000005F



overall
accuracy:
96.72%

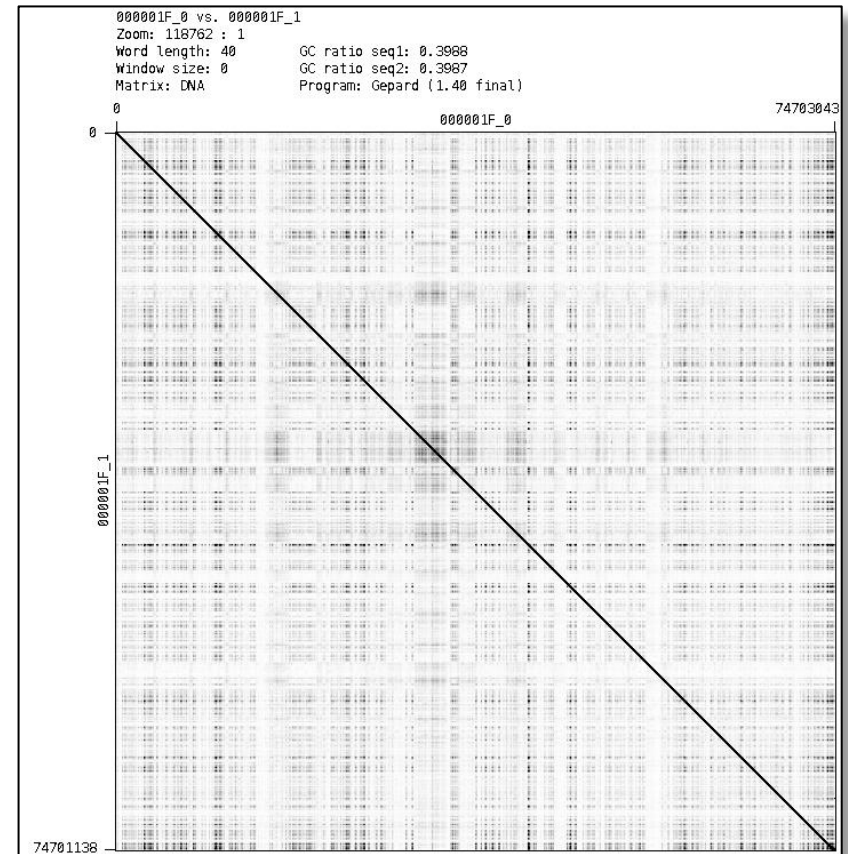
APPLICATION TO HUMAN ASSEMBLY

FALCON-Unzip



- X-axis – 75Mb contig
- Y-axis – Hundreds of haplotigs spanning 87% of the primary contig

FALCON-Phase



- X-axis – 75Mb contig – Phase 0
- Y-axis – 75Mb contig – Phase 1